Molecular characteristics of purinergic receptors and its mediated signaling, a novel target for stem cell-based therapy: A brief summary

Anbuselvi Mattuvar Kuzhali S

Department of Physiology, Institute of Physiology and Experimental Medicine, Madras Medical College, Chennai, Affiliated to the Tamil Nadu Dr. MGR Medical University, Chennai, Tamil Nadu, India

Abstract

Molecular identity of nucleotide factor(s) that serve as trigger for the activation of cell-cycle progression in adult stem cells or human mesenchymal stem cells (hMSCs) or progenitor cells is not well understood. Recent studies highlight the importance of extracellular nucleotide-triphosphates (eNTPs) like ATP as key regulators of various functions executed by the adult stem cells. It has long been speculated that discrete release of ATP by the cells in response to shear stress plays a major role in the initiation of various physiological processes such as proliferation, repair and regeneration, migration and homing. eNTPs acts via purinoceptors (P2Y) and multiple isoforms of P2Y receptor are expressed in all the cells, the molecular mechanisms of P2Y-signaling still remain unexplored. On certain stimuli, hMSCs release ATP constitutively and ATP activates variety of P2X/P2Y receptors in MSCs inducing robust Ca²⁺ responses from different tissues/species and calcium mediated functional responses in hMSCs were considered as predominant one. The mechanism of store-operated Ca²⁺ entry through which the intracellular calcium concentration is restored plays a major role in that. Yet the cognate intrinsic mechanisms of these purinergic receptors remain contentious. Our interest is to understand the basic mechanisms under which the stem cells perform their functions by strengthening the signaling cascades and genetic transcription effected by activation of purinoceptors.

Keywords: ATP receptors, calcium signalling, human mesenchymal stem cells

Corresponding Author

Dr. Anbuselvi Mattuvar Kuzhali S, Assistant Professor, Department of Physiology, Institute of Physiology and Experimental Medicine, Madras Medical College, Chennai. Telephone: + 9940804760, Email: dranbuselvimk@gmail.com

Introduction

Extracellular ATP related nucleotides behave as trophic, immunomodulatory molecules through its purinergic receptors: P1 receptors for nucleosides and P2 receptors for nucleotides. Purinergic ligands are well recognized mediators of autocrine/paracrine signaling involved in proliferation, migration, immunomodulation and regeneration.¹ An upcoming interest in translational research is on cell based therapieswith human mesenchymal stem cells (hMSCs), a cell-population with self-renewal and

multipotency that was derived initially from the bone marrow, later from the adipose tissues, Wharton's jelly/placenta and dental pulp.²⁻⁸ The isolated cells are recognized as hMSCs only after confirming their morphology and functional characteristics by series of tests that specifically proves their adherence ability, positive expression for specific surface markers(CD), the clonogenic assays and differentiating abilities maintaining their lineage properties.^{5,9,10} This review briefly discuss the recent advances in molecular characterization of purinergic receptors, their established novel physiological

processes and how they act as ideal candidates to mediate the crosstalk with resident stem cell niches, promoting cell growth and survival, differentiation, immunomodulation, migration and repair.

Extracellular Purines - A Heterodox Concept

From an evolutionary point of view, the most ancient recognized molecules with autocrine/paracrine signaling are the extracellular nucleotides (eNTPs). They are released during stress/hypoxia and comprises of both extracellular purines (ATP, ADP, β-NAD. ADPR and cADPR) and extracellular pyrimidines (UTP and UDP) in the extracellular mileu.¹ Most recent research is being highly focused on the role of purine nucleotide ATP (enzyme cofactor). In 1929, ATP was first isolated by Karl Lohmann and documented as signaling molecule by Drury andSzent.¹¹⁻¹³ Only decades later, ATP was accepted as potent extracellular messengers producing effects on energy metabolism, signal transduction and cellular communication.¹⁴⁻¹⁶ The release of eNTPs is regulated by leakage through ATP-binding cassette transporters, connexin hemichannels, or voltagedependent anion channels or by receptor mediated exocytosis.¹⁷ It was proven that eNTPs bind a distinct cell surface receptors, the Purinergic receptor family.¹³

Purinergic receptors

Purinergic receptors, the oldest receptors, were initially reported in gut smooth muscles in 1970s.¹⁸⁻²⁰ Purinoceptors have been proved to have role in cell proliferation, differentiation and migration in endothelial cells, tumour cells, embryonic or adult stem cells.²¹⁻²³ Based on the order of activation of agonists, agonists efficacy and desensitization characteristics to its antagonists the purinoceptors were classified into P1 receptors (adenosine receptors) with its natural ligand adenosine and P2 receptors(nucleotide receptors) activated by other nucleotides(ATP, UTP).²⁴ The seven trans-membrane domain P1 receptors comprises of A1, A2A, A2B and A3, mediates its effects via phospholipase C, adenylyl cyclaseand mitogen-activated protein kinase pathway.^{25,26}

Ectoenzymes: Currently, an accumulating evidence suggests that a family of ectonucleotidases (E-NTPases or ectonucleoside triphosphate diphosphohydrolases) called ectoenzymes on the

plasma membrane, play a role in the nucleotide metabolism.²⁷⁻²⁹ Once released, ATP mediates signaling via purinoceptors or degraded to nucleosides (AMP, adenosine) by the sequential of four ectoenzymes: ectonucleotide action pyrophosphatase, ectonucleotide triphosphate diphosphohydrolase/CD39 (E-NTPDase), alkaline phosphatase, and ecto-5'-nucleotidase/CD73.³⁰⁻³³

P2 receptors exist as two distinct families: The metabotropic P2Y and ionotropic P2X receptorsare widely distributed on both excitable and nonexcitable mammalian cells. P2Xare cationselective, ligand-gated ion channels (LGIC) on the plasma membrane activated by ATP and P2Y are Gprotein coupled receptors activated by nucleotides, di-phosphates or triphosphates, purines and pyrimidines.^{34,35} However, several studies states that seven out of eight members of the P2X receptor family (P2X1-7) expressed in splice variants and approximately six out of 11 types of P2Y receptors (P2Y1, P2Y2, P2Y4, P2Y6, P2Y11, and P2Y12) expressed were documented to have functional responses.¹³ P2 and P1 receptors mediate varied signaling events, where the ligand and receptor affinity of P2 and P1 differs from nanomolar to micromolar levels.^{36,37}

- P2=affinity range from nanomolar (e.g., P2Y2) to micromolar (e.g., P2X7) levels;
- P1=affinity range from low nanomolar (e.g., A1,A2A,A3) to high nanomolar (e.g., A2B) levels.

P2X and P2Y receptors: Purinergic signaling is also modulated by cross-talk between P1 and P2 receptors.³⁸ Studies have well documented the role of ATP via P2R in hMSC differentiation.³⁹⁻⁴¹

- P2Y receptors in the adipogenic/osteogenic differentiation of hMSCs.⁴²
- P2X receptors on proliferation(P2X1/5) and differentiation of hMSCs (P2X5 and P2X7).²³

Of the seven different subunits (P2X1-7)P2X1-5 are homomeric receptors and P2X2/3 and P2X1/5 receptors were heterodimer, with the exception of P2X6 and P2X7.⁴³⁻⁴⁶Functional responses P2X receptors are mediated by increasing intracellular calcium ion concentration, yet P2X7 was proven to mediate via MAP-kinase, heat shock proteins, and 4-kinase.47-50 The phosphatidylinositol P2Y metabotropic, heptahelical G-protein coupled receptor (GPCR) family has eight subtypes in human tissues-P2Y1, P2Y2, P2Y4, P2Y6, P2Y11, P2Y12, P2Y13 and P2Y14 and are activated by nucleotides like

ADP,ATP, UDP and UTP.⁵¹ They couple to Gq and activate phospholipase C- β (P2Y-1,2,4,6,11), or to Gi(P2Y-12,14)that inhibit adenylyl cyclase whereas P2Y11 are coupled to both Gq/Gsand increase intracellular Ca2 and cAMP levels.⁵²

Purinoceptors in MSCs

MSCs are constituitively activated by ATP upon stimuli and the presence of purinergic receptors and ectoenzymes on their plasma membrane.⁵³⁻⁵⁵ Evans and coworkers were the first who evidenced the presence of P1 receptor in stem cell surface.⁵⁶ Both invitro and invivo studies have put forth the role of P1 receptors in the process of adipogenic differentiation (A1,A2A), osteogenic differentiation (A2B) and cytokine activation in Bone marrow MSCs by gene knockout studies as well as pharmacological deletion/blockade of P1 receptors.⁵⁷⁻⁶⁴

Advances in the molecular biology are used to identify novel ligands for P2 receptors and its relation to hMSC has gained attention only recently. Studies states that the spontaneous release of ATP from MSCs occurs via pannexins hemichannels (gap junction) on the plasma membrane and its direct stimulation of P2Y1 receptor was associated with increase in intracellular Ca²⁺levels.⁵⁵ This study hypothesized that modulation of the proliferation rate of MSCs at early passages cultures could be achieved by the concurrent activation of P2X and P2Y receptors by ATP analogs/agonists.²³ A gene knockout study in rat MSCs suggested that the activation of P2Y2 receptor induced intracellular Ca2+ responses that directly correlated with cell cycle progression.⁶⁵ Currently, an in-vivo study by Ferrari and collaborators observed an upregulation of proinflammatory cytokines and cell migration related genes in ATP pretreated MSCs, enhanced the migratory and homing ability of hMSCs, this concept was considered supportive in therapies for bone marrow engraftment.66

Plausible purinergic cascade in hMSCs^{22,39,41,43,55}

 eNTPs increases the intracellular cytosolic Ca²⁺ levels ([Ca²⁺]i) in hMSCs via enhancing Ca²⁺ influx through plasma-membrane-calciumchannels(PMCA) and hemichannels (pannexins) as well as Ca²⁺release from intracellular stores of mitochondria and endoplasmic reticulum (ER).

- eNTPs binds to G-protein-coupled-P2Y receptors, can elevate the [Ca²⁺]i by stimulating phospholipase-C to generate inositol-1,4,5-triphosphate(IP3), which in-turn activates the IP3 receptor on endoplasmic reticulum (ER) and induces Ca²⁺ release from ER.
- Thus the reduction Ca²⁺ level inside ER activates the Stim1 channels on ER-membrane which aggregates and coactively binds to ORAI-1 channels in plasma membrane causing increased Ca²⁺ influx-intracellularly. This mechanism called Ca²⁺ release-activated Ca²⁺ channels (CRAC) enables reconstituition of cytosolic Ca²⁺ levels. STIM1 and ORAI-1 are the essential components of CRAC channels that mediate store-operated²⁺ Ca²⁺entry (SOCE) in cells.
- Increases in the concentration of either calcium induce cAMP pathway that will further trigger the activation of kinasesa and cAMP responsive element binding protein, a nuclear protein that modulates the transcription of genes for chemotaxis, cellular motility, proliferation, immunomodulation and repair.

Methods of study 58,66-69

Expression of purinoceptors, ion channel and transduction pathways in hMSCs can be studied by Western blot method and qPCR. Immunocytochemistry will help in studying the underlying cytokines. Recording of intracellular calcium concentration and functioning of calcium ion channels can be studied by single cell imaging using fura-2 based radiometry. This method will clearly delineate the mechanism of ER-Ca⁺stores depletion by using specific and nonspecific Calcium-ATPase blockers cyclopiazonic acid (CPA) or thapsigargin(TG) to Orai1/Stim1channels. Animal models by method of knockout-mice (gene specific for the targeted receptor is deficient or deleted) has been the best proven method to assess the functionale of purinoceptors in the absence of selective antagonists or antisense probes.

P2 receptors and ectoenzymes are dynamic cellular entities. Studies have proved that P2Y receptors and ecto-59-nucleotidase, ecto-apyrase and E-NTPaseundergo stage-specific transient expression during cellular movement.⁶⁷ Studies are becoming increasingly evident on the role of altered E-NTPase activities as novel drug targets. Apart from ligand binding assays, functional fluorescent imaging (FLIPR) in cell lines transfected animal models has been suggested the advanced screening approach to identify newer ligands to human P2 receptors.^{68,69}

Conclusion

Researchers have their current interests on identifying the crosstalk between biochemical signals produced by MSCs in tissue environments. However, some have revealed unexpected findings about eNTPs and their derivatives in modulating the physiology of stem cells. From what has been discovered so far, a brief insight about the purinergic receptors has been summarized. We hope that this will help researchers to precisely define these molecular signals focusing their importance for future hMSC-based therapies.

Acknowledgement: Nil

Conflicts of Interest: Nil

References

- Schwiebert EM, Fitz JG. Purinergic signaling microenvironments: An introduction. Purinergic Signal. 2008; 4:89-92.
- 2. Salem HK, Thiemermann C. Mesenchymal stromal cells: current understanding and clinical status. Stem Cells.2010; 28:585-96.
- English K. Mechanisms of mesenchymal stromal cell immunomodulation. Immunol Cell Biol. 2013; 91:19-26.
- Friedenstein AJ, Chailakhjan RK, Lalykina KS. The development of fibroblast colonies in monolayer cultures of guinea pig bone marrow and spleen cells. Cell Tissue Kinet.1970; 3:393-403.
- Pittenger MF, Mackay AM, Beck SC, Jaiswal RK, Douglas R, Mosca JD, Moorman MA, Simonetti DW, Craig S, Marshak DR. Multilineage potential of adult human mesenchymal stem cells. Science.1999; 284:143-47.

- Mosna F, Sensebé L, Krampera M. Human bone marrow and adipose tissue mesenchymal stem cells: a user's guide. Stem Cells Dev. 2010; 19:1449-70.
- Batsali AK, Kastrinaki MC, Papadaki HA, Pontikoglou C. Mesenchymal stem cells derived from Wharton's Jelly of the umbilical cord: biological properties and emerging clinical applications. Curr Stem Cell Res Ther.2013; 8:144-55.
- Huang GT, Gronthos S, Shi S. Mesenchymal stem cells derived from dental tissues vs. those from other sources: their biology and role in regenerative medicine. J Dent Res. 2009; 88:792-806.
- 9. Wagner W, Ho AD. Mesenchymal stem cell preparations- comparing apples and oranges. Stem Cell Rev. 2007; 3:239-48.
- 10. Ho AD, Wagner W, Franke W. Heterogeneity of mesenchymal stromal cell preparations. Cytotherapy.2008; 10:320-30.
- 11. LohmannK.Uberdiepyrophosphatfraktionimmuskel.Naturwissenschaften.1929; 17:624–5.
- Drury AN, Szent-Gyeorgyi A. The physiological activity of adenine compounds with especial reference to their action upon the mammalian heart. J Physiol. 1929; 68:213–37.
- 13. Ralevic V, Burnstock G. Receptors for purines and pyrimidines. Pharmacol Rev. 1998; 50:413–92.
- Berne RM, Rubio R. Adenine nucleotide metabolism in the heart. Circ Res. 1974; 35[Suppl 3]:109–20.
- 15. Fredholm BB. Adenosine, an endogenous distress signal, modulates tissue damage and repair. Cell Death Differ. 2007; 14:1315–23.
- 16. Burnstock G. Pathophysiology and therapeutic potential of purinergic signaling. Pharmacol Rev.2006; 58:58–86.
- 17. Burnstock G. Historical review: ATP as a neurotransmitter. Trends Pharmacol Sci. 2006; 27:166-176.
- Burnstock G, Verkhratsky A, et al. Evolutionary origins of the purinergicsignalling system. Acta Physiol. 2009; 195:415–47.
- 19. Burnstock G. Purinergic nerves. Pharmacol Rev. 1972; 24:509–81.
- 20. Burnstock G. A basis for distinguishing two types of purinergic receptor. In: Straub RW, Bolis L, eds. Cell membrane receptors for

drugs and hormones: a multidisciplinary approach. New York: Raven Press; 1978. p. 107–18.

- Evans BA, Elford C, Pexa A, Francis K, Hughes AC, Deussen A. Human osteoblast precursors produce extracellular adenosine, which modulates their secretion of IL-6 and osteoprotegerin. J Bone Miner Res. 2006; 21:228–36.
- Kawano S, Otsu K, Kuruma A, Shoji S, YanagidaE,Muto Y. ATP autocrine/paracrine signaling induces calcium oscillations and NFAT activation in human mesenchymal stem cells. Cell Calcium.2006; 39:313–24.
- 23. Coppi E, Pugliese AM, Urbani S, Melani A, Cerbai E, Mazzanti B. ATP modulates cell proliferation and elicits two different electrophysiological responses in human mesenchymal stem cells. Stem Cells.2007; 25:1840–9.
- Fredholm BB, IJzerman AP, Jacobson KA, Linden J, Müller CE. International Union of Basic and Clinical Pharmacology. LXXXI. Nomenclature and classification of adenosine receptors—an update. Pharmacol Rev. 2011; 63:1-34.
- Burnstock G. Purinergic signaling and vascular cell proliferation and death. ArteriosclerThrombVasc Biol. 2002; 22:364– 73.
- Gessi S, Merighi S, Sacchetto V, Simioni C, Borea PA. Adenosine receptors and cancer. BiochimBiophysActa. 2011; 1808:1400–12.
- Zimmerman H. Two novel families of ectonucleotidases: Molecular structures, catalytic properties and a search for function. Trends Pharmacol Sci. 1999a; 20:231-36.
- 28. Goding JW, Howard MC. Ecto-enzymes of lymphoid cells. Immunol Rev. 1998; 161:5-10.
- 29. Burnstock G. Release of vasoactive substances from endothelial cells by shear stress and purinergicmechano-sensory transduction. J Anat. 1999; 194:335-42.
- Rossi L, Salvestrini V, Ferrari D, Di Virgilio F, Lemoli RM. The sixth sense: hematopoietic stem cells detect danger through purinergic signaling. Blood.2012; 120:2365-75.
- Zimmermann H. Extracellular metabolism of ATP and other nucleotides. NaunynSchmiedebergs Arch Pharmacol.2000; 362: 299-309.
- 32. Latini S, Pedata F. Adenosine in the central nervous system: release mechanisms and

extracellular concentrations. J Neurochem.2001; 79:463-84.

- 33. Heinrich A, Ando RD, Túri G, Rozsa B, Sperlagh B. K+ depolarization evokes ATP, adenosine and glutamate release from glia in rat hippocampus: a microelectrode biosensor study. Br J Pharmacol. 2012; 167:1003-20
- 34. Von Keugelgen I, Harden TK. Molecular pharmacology, physiology and structure of the P2Y receptors. AdvPharmacol. 2011; 61:373-415.
- Boeynaems JM, Communi D, Gonzalez NS, Robaye B. Overview of the P2 receptors.SeminThrombHemost. 2005; 31:139–49.
- 36. Fredholm BB. Adenosine, an endogenous distress signal, modulates tissue damage and repair. Cell Death Differ. 2007; 14:1315–23.
- 37. Junger WG. Immune cell regulation by autocrinepurinergicsignalling. Nat Rev Immunol.2011; 11:201–12.
- Sichardt K, Nieber K. Adenosine A1 receptor: Functional receptor-receptor interactions in the brain. Purinergic Signal. 2007; 3:285-98.
- 39. Sun D, Junger WG, Yuan C, Zhang W, Bao Y, Qin D, Wang C, Tan L, Qi B, Zhu D, Zhang X, Yu T. Shockwaves induce osteogenic differentiation of human mesenchymal stem cells through ATP release and activation of P2X7 receptors. Stem Cells.2013; 31:1170-80.
- 40. Ciciarello M, Zini R, Rossi L, Salvestrini V, Ferrari D, Manfredini R, Lemoli RM. Extracellular purines promote the differentiation of human bone marrowderived mesenchymal stem cells to the osteogenic and adipogenic lineages. Stem Cells Dev. 2013; 22:1097-1111.
- 41. Biver G, Wang N, Gartland A, Orriss I, Arnett TR, Boeynaems JM, Robaye B. Role of the P2Y13 receptor in the differentiation of bone marrow stromal cells into osteoblasts and adipocytes. Stem Cells.2013; 31:2747-58.
- 42. Zhang Y, Lau P, Pansky A, KassackM, Hemmersbach R, Tobiasch E. The influence of simulated microgravity on purinergic signaling is different between individual culture and endothelial and smooth muscle cell coculture. Biomed Res Int. 2014; 2014article ID: 413708.
- Coddou C, Yan Z, Obsil T, Huidobro-Toro JP, Stojilkovic SS. Activation and regulation of purinergic P2X receptor channels. Pharmacol Rev. 2011; 63:641–83.

- 44. Compan V, Ulmann L, Stelmashenko O, Chemin J, Chaumont S, Rassendren F. P2X2 and P2X5 subunits define a new heteromeric receptor with P2X7-like properties. J Neurosci.2012; 32:4284–96.
- 45. Torres GE, Egan TM, Voigt MM. Heterooligomeric assembly of P2X receptor subunits. Specificities exist with regard to possible partners. J Biol Chem. 1999; 274:6653–59.
- 46. Barrera NP, Ormond SJ, Henderson RM, Murrell-Lagnado RD, Edwardson JM. Atomic force microscopy imaging demonstrates that P2X2 receptors are trimmers but that P2X6 receptor subunits do not oligomerize. J Biol Chem. 2005; 280:10759–65.
- 47. Kim M, Jiang LH, Wilson HL, North RA, Surprenant A. Proteomic and functional evidence for a P2X7 receptor signalling complex. EMBO J. 2001; 20:6347–58.
- Ghiringhelli F, Apetoh L, Tesniere A, Aymeric L, Ma Y, Ortiz C. Activation of the NLRP3 inflammasome in dendritic cells induces IL-1beta-dependent adaptive immunity against tumors. Nat Med 2009; 15:1170–78.
- 49. Yegutkin GG. Nucleotide and nucleosideconverting ectoenzymes: important modulators of purinergicsignalling cascade. BiochimBiophysActa. 2008; 1783:673–94.
- Pellegatti P, Falzoni S, Pinton P, Rizzuto R, Di Virgilio F. A novel recombinant plasma membrane-targeted luciferase reveals a new pathway for ATP secretion. MolBiol Cell.2005; 16:3659-65.
- 51. Abbracchio MP, Burnstock G, Boeynaems JM, Barnard EA, Boyer JL, Kenedy C. International Union of Pharmacology LVIII: update on the P2Y G protein-coupled nucleotide receptors: from molecular mechanisms and pathophysiology to therapy. Pharmacol Rev. 2006; 58:281-341.
- 52. Boeynaems JM, Communi D, Gonzalez NS, Robaye B. Overview of the P2 receptors. SeminThrombHemost. 2005; 31:139-49.
- 53. Fruscione F, Scarfi S, Ferraris C, Bruzzone S, Benvenuto F, Guida L, Uccelli A, Salis A, Usai C, Jacchetti E, Ilengo C, Scaglione S, Quarto R, Zocchi E, De Flora A. Regulation of human mesenchymal stem cell functions by an autocrine loop involving NAD+ release and P2Y11-mediated signaling. Stem Cells Dev. 2011; 20:1183-98.

- 54. Chen M, Su W, Lin X, Guo Z, Wang J, Zhang Q, Brand D, Ryffel B, Huang J, Liu Z, He X, Le AD, Zheng SG. Adoptive transfer of human gingiva derived mesenchymal stem cells ameliorates collagen-induced arthritis via suppression of Th1 and Th17 cells and enhancement of regulatory T cell differentiation. Arthritis Rheum.2013; 65:1181-93.
- 55. Kawano S, Otsu K, Kuruma A, Shoji S, Yanagida E, Muto Y, Yoshikawa F, Hirayama Y, Mikoshiba K, Furuichi T. ATP autocrine/paracrine signaling induces calcium oscillations and NFAT activation in human mesenchymal stem cells. Cell Calcium.2006; 39:313-24.
- 56. Evans BA, Elford C, Pexa A, Francis K, Hughes AC, Deussen A, Ham J. Human osteoblast precursors produce extracellular adenosine, which modulates their secretion of IL-6 and osteoprotegerin. J Bone Miner Res. 2006; 21:228-36.
- 57. Napieralski R, Kempkes B, Gutensohn W. Evidence for coordinated induction and repression of ecto-5'-nucleotidase (CD73) and the A2a adenosine receptor in a human B cell line. Biol Chem. 2003; 384:483-87.
- 58. Gharibi B, Abraham AA, Ham J, Evans BA. Adenosine receptor subtype expression and activation influence the differentiation of mesenchymal stem cells to osteoblasts and adipocytes. J Bone Miner Res. 2011; 26:2112-24.
- 59. Carroll SH, Wigner NA, Kulkarni N, Johnston-Cox H, Gerstenfeld LC, Ravid K. A2B adenosine receptor promotes mesenchymal stem cell differentiation to osteoblasts and bone formation in vivo. J Biol Chem. 2012; 287:15718-27.
- 60. He W, Mazumder A, Wilder T, Cronstein BN. Adenosine regulates bone metabolism via A1, A2A, and A2B receptors in bone marrow cells from normal humans and patients with multiple myeloma. FASEB J. 2013; 27:3446-454.
- 61. Fredholm BB. Adenosine receptors as drug targets. Exp Cell Res. 2010; 316:1284-88.
- 62. Katebi M, Soleimani M, Cronstein BN. Adenosine A2A receptors play an active role in mouse bone marrow-derived mesenchymal stem cell development. J Leukoc Biol. 2009; 85:438-44.

- 63. Sun LL, Xu LL, Nielsen TB, Rhee P, Burris D. Cyclopentyl adenosine improves cell proliferation, wound healing and hair growth. J Surg Res. 1999; 87:14-24.
- 64. Shimegi S. ATP and adenosine act as a mitogen for osteoblast-like cells (MC3T3-E1). Calcif Tissue Int. 1996; 58:109-13.
- Ichikawa J, Gemba H. Cell density-dependent changes in intracellular Ca²⁺ mobilization via the P2Y2 receptor in rat bone marrow stromal cells. J Cell Physiol. 2009; 219:372-81.
- 66. Ferrari D, Gulinelli S, Salvestrini V, Lucchetti G, Zini R, Manfredini R, Caione L, Piacibello W, Ciciarello M, Rossi L, Idzko M, Ferrari S, Di Virgilio F, Lemoli RM. Purinergic stimulation of human mesenchymal stem cells potentiates their chemotactic response to CXCL12 and increases the homing capacity and production of proinflammatory cytokines. ExpHematol. 2011; 39:360-74, 374.e1-5.
- 67. Dubyak GR, Clifford EE, Humphreys BD, Kertsey SB, Martin KA. Expression of multiple ATP subtypes during the differentiation and inflammatory activation of myeloid leukocytes. Drug Dev Res. 1996; 39:269–78.
- Bianchi BR, Lynch KJ, Touma E, Niforatos W, Burgard EC, Alexander KM, Park HS, Yu H, Metzger R, Kowaluk E, Jarvis MF, van Biesen T. Pharmacological characterization of recombinant human and rat P2X receptor subtypes. Eur J Pharmacol. 1999; 376:127-38.
- **69.** Williams M, Jarvis MF. Purinergic and pyrimidinergic receptors as potential drug targets. Biochem Pharmacol. 2000; 59:1173-85.